AMENDMENTS TO THE SPECIFICATION

Replace the paragraph at page 30, lines 8-29, with:

<Example 2> Cloning and expression of DnaK fragments

A DnaK fragment was cloned by amplifying an objective gene fragment using PCR method with genomic DNA extracted from *Escherichia coli* K-12 strain as a template. KOD-Plus- supplied from Toyobo Co., Ltd. was used for PCR amplification of the gene. Concretely, in the amplification of DnaK 384-638, a sample was prepared to contain reaction buffer, 1 mM $MgSO_4$, 15 pmole primers shown in SEQ ID NOS:3 and 4, 1 unit of polymerase and 100 ng of *Escherichia coli* DNA in 50 μL of a reaction solution, and subjected to the reaction of 25 cycles at 94°C for 15 seconds, 55°C for 30 seconds and 68°C for one minute after the reaction at 94°C for 2 minutes. ~~Likewise, the primers shown in SEQ ID NOS:3 and 4 were used for the amplification of DnaK 386-586.~~ An amplified DNA fragment was digested with a restriction enzyme BamHI, and cloned into a BamHI-SmaI site of pQE30 (the DNA fragment amplified using KOD-Plus-(supplied from TOYOBO) had a blunt end, and thus a downstream side of the amplified fragment was directly used). The sequence of the cloned gene was confirmed by sequencing analysis. A 6xHis sequence (histidine tag) can be added to the N terminus of the objective protein by cloning into this vector. An expression plasmid, pQE-DnaK 384-638 was made in this way.

Replace the paragraph at page 31, lines 24-34, with:

<Example 4> Preparation of N terminus-deleted DnaK clone

The PCR amplification was performed using pQE-DnaK 384-607 produced in Example 2 as the template and the primers shown below, and an amplified fragment was introduced into the BamHI-SmaI site of the pQE30 vector in accordance with the method in Example 2. The produced clones and sets of the primers are as follows. DnaK 508-607: SEQ ID NOS:13 and ~~2~~ 4, DnaK 525-607: SEQ ID NOS:14 and ~~2~~ 4, and DnaK 419-607: SEQ ID NOS:15 and ~~2~~ 4. The sequence of each mutant produced was confirmed by sequencing analysis, and subsequently the protein was purified in accordance with the method described in Example 2.